

# Determining Plasma Protein Binding of Liraglutide with the new *EScalate* Equilibrium Shift Assay

J. Ungewiss<sup>1</sup>, S. Gericke<sup>1</sup>, H. Boriss<sup>2</sup>

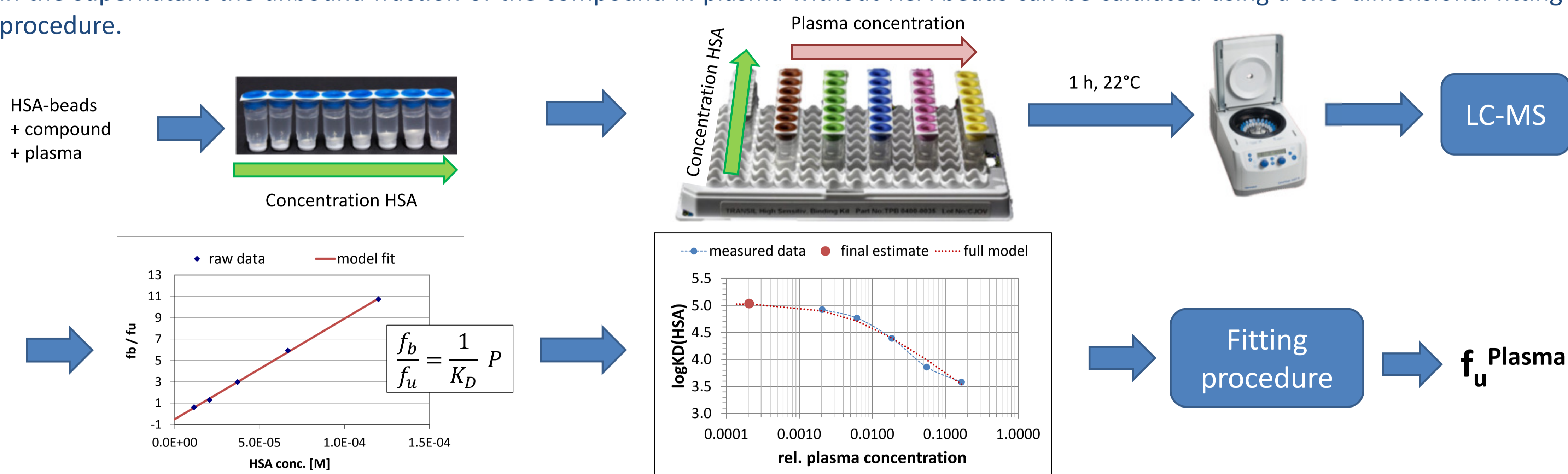
<sup>1</sup>3B Pharmaceuticals GmbH, Berlin, Germany; <sup>2</sup>Sovicell GmbH, Leipzig, Germany

## Background

The accurate determination of plasma protein binding of larger molecules represents a significant challenge, especially if their plasma protein binding is very high. The commonly used equilibrium dialysis and ultrafiltration methods are often not applicable for peptides and oligonucleotides due to low migration velocities through the semipermeable membrane. Moreover, substantial non-specific binding to the membrane and limited solubility in protein-free aqueous buffer further limit the application of those assays. In order to overcome these limitations, we developed the bead-based *EScalate* equilibrium shift *in vitro* assay. The applicability of the assay was shown for the peptidic drug liraglutide and the small molecule drugs carbamazepine, desipramine, pyrimethamine and warfarin.

## *EScalate* assay principle

The *EScalate* assay is based on the determination of the binding of the compound to HSA immobilized on beads. By adding plasma to the supernatant over the HSA beads the compound also binds to plasma proteins. The equilibrium of the bead binding is shifted resulting in a lower apparent  $K_D$  for the bead binding. By independent variation of both the amount of HSA coated beads and the plasma concentration in the supernatant the unbound fraction of the compound in plasma without HSA beads can be calculated using a two-dimensional fitting procedure.



## Results

First, the accuracy of the *EScale* assay was confirmed with the four well-studied small molecule drugs, carbamazepine, desipramine, pyrimethamine and warfarin (table 1).

Then, the plasma protein binding data for the high serum albumin binding peptidic drug liraglutide was determined five-fold on 3 different days (table 2).

**Table 1:** Comparison of the unbound fraction  $f_u$  in human plasma determined by *EScale* with data obtained by ultrafiltration (UF) oder equilibrium dialysis (ED)

Compound	$f_u$ <i>EScale</i>	$f_u$ Literature
Carbamazepine	28.8%	28.0% (UF) <sup>1</sup>
Desipramine	17.3%	14.0% (ED) <sup>2</sup>
Pyrimethamine	12.5%	11.7% (ED) <sup>3</sup>
Warfarin	0.55%	0.69% (ED) <sup>4</sup>

**Table 2:** Unbound fraction  $f_u$  of Liraglutide in human plasma determined by *EScale*

Measurement	$f_u$ Liraglutide
Day 1 run 1	0.66%
Day 1 run 2	0.58%
Day 2 run 1	0.40%
Day 2 run 2	0.44%
Day 3 run 1	0.47%
<b>Average</b>	<b>0.51%</b>
Literature <sup>5</sup>	0.53%

*Advantages of the ESscale assay:*

- **No dialysis membrane** used
- **Rapid equilibration** at room temperature
- **Presence of plasma in all samples** to prevent non-specific binding to vessels and compound precipitation
- **Capability to analyze strong plasma protein binders** due to indirect determination of  $f_u^{\text{Plasma}}$

Literature:

1. Fortuna A, Alves G, Soares-da-Silva P, Falcão A 2013. *Epilepsy Research* 107(1):37-50.
2. Zhang F, Xue J, Shao J, Jia L 2012. *Drug Discovery Today* 17(9-10):475-485
3. Rudy AC, Poynor WJ 1990. *Pharmaceutical Research* 7(10):1055-1060.
4. Mungall D, Wong YY, Talbert RL, Crawford MH, Marshall J, Hawkins DW, Ludden TM 1984. *Journal of Pharmaceutical Sciences* 73(7):1000-1001.
5. Flint, A.; Nazzari, K.; Jagielski, P.; Hindsberger, C.; Zdravkovic, M., *British Journal of Clinical Pharmacology* 2010, 70 (6), 807-814.

## Conclusion

The newly developed *EScale* assay is providing accurate results across a broad range of plasma protein binding levels. It is unique in providing reliable data for mid-sized organic molecules with high affinity to plasma proteins.